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ANALOGS OF THE ORALLY ACTIVE GROWTH HORMONE SECRETAGOGUE L-162,752

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Abstract: A series of spiroindane growth hormone secretagogues that vary in their amino side chains is reported. Variations in these side chains markedly affect growth hormone release *in vitro* and *in vivo*. The best side chain in this series of secretagogues is α-methylalanine. L-162,752 not only stimulated GH release from rat pituitary cells, but also induced GH release in dogs upon i.v. and p.o. administration.

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Recombinant growth hormone (GH) is currently used clinically to improve the growth of GH deficient children and to treat GH deficient adults and patients with Turner's syndrome. A number of additional uses are understudy which derive from its anabolic properties. However, the use of GH has limitations because it is a relatively large polypeptide (191 amino acids) which must be administered by injection. An alternative is to stimulate release of GH from the pituitary gland. Recently both peptide GH secretagogues including GHRP-6^{1a} and hexarelin^{1b} and a nonpeptide GH secretagogue L-692,429² have shown efficacy and specificity in the release of GH in clinical trials. Unfortunately these secretagogues have poor oral bioavailability. More recently our group has reported a highly potent, orally active GH secretagogue L-163,191 (MK-0677) which has been selected for clinical studies.³ In this communication, we report the evaluation of a variety of amino side chains which formed the basis for using the aminoisobutyric acid side chain in L-163,191.

The design of our orally active secretagogues originated in a project to derivatize "privileged structures" as a strategy to discover leads for G-protein-linked receptors.³ The lead compound, a spiroindanylpiperidine 1, stimulated GH release in our rat pituitary cell assay⁴ in a dose-dependent manner with an EC₅₀ value of 50 nM. This intrinsic potency is especially remarkable since 1 is a mixture of four diastereomers. Compound 1 when given intravenously at a dose of 0.1 mg/kg to Beagle dogs⁵ increased serum GH levels, however, it was poorly active when administrated orally.⁶

An extensive program was undertaken to replace the quinuclidinyl urea of 1 on the assumption that its polarity and strongly basic properties might hinder oral absorption. The highlight of that effort was the replacement of the quinuclidine urea of 1 with an α -methylalanine moiety with both (R)- and (S)- tryptophan to afford the individual antipodes 2 (L-162,752) and 3. The more active of these compounds L-162,752 was the first orally active GH secretagogue to be synthesized from the spiroindanylpiperidine lead 1 (see below). Herein we report the preparation and biological activities of L-162,752 and its related analogs.

Chemistry

The synthesis of GH secretagogue analogs is illustrated in Schemes 1 and 2 and involved the coupling of Cbz or Boc protected D-tryptophan to the spiroindanylpiperidine 48 using EDC and HOBt under standard peptide coupling conditions. The protecting group was removed to give 5 which served as a convenient precursor for a variety of structurally diverse amino side chains which were added via a second peptide-type coupling reaction followed by a final deprotection of the coupled product I with HCl/Et₂O.

N-substituted derivatives II were prepared by a reductive amination of 2 and 6 with the appropriate aldehyde (Scheme II). For example, the synthesis of the hydroxypropyl derivatives 20 and 22 was accomplished using (R)-2-tetrahydropyranyloxypropanal 18¹⁰ and the dihydroxypropyl derivative 21 was prepared from (R)-2,3-O-

isopropylidene-glyceraldehyde 19.11 The GH secretagogues 20, 21 and 22 were isolated after removing the tetrahydropyranyl or acetonide groups under acidic conditions.

Results and Discussion

Growth hormone release in vitro was determined in rat pituitary cell assays.⁴ Development of the lead compound 1 initially focused on the introduction of amino side chains which were known to work well in the benzolactam GH secretagogues.⁷ One of the simplest and most active of these is aminoisobutyric acid and when it was used with (R)- and (S)-tryptophan the high activity of L-162,752 established the importance of (R)- stereochemistry in the central amino acid. It is interesting that the more active benzolactam secretagogues also have an (R)stereochemistry whose modeling suggested coincidence with the (D)-Trp of GHRP-6.7 Subsequent analogs which are reported herein were all synthesized as (D)-Trp derivatives. Presented in Table 1 are results with some of the analogs that helped define the optimal amino acid side chain of these spiroindanylpiperidine GH secretagogues. Systematic modification of the α-methylalanine side chain confirmed the dependence of GH secretagogue activity on both chain length and substitution pattern. The homologated analog 6 shows a 5-fold loss in potency compared with 2. The importance of the geminal dimethyl substitution flanking the basic amine is reflected in the poor activities of the alanine derivatives 7 and 8 (both D- and L-), and of the glycine derivative 9. The spiro substituted glycine series (analogs 10-12) revealed several important trends. Substitution with spirocyclopentane (analog 11) and spirocyclohexane (analog 12) groups reduces intrinsic activity significantly. These results suggest limited space in the receptor binding site adjacent to the carbonyl group. Evident from the poor activity of analog 10 (pKa 5.9)¹² is the requirement of a basic amine in achieving good GH secretagogue activity. ¹³ Comparison of the activities of the (S)- α -methylserine derivative 13 with that of the (R)- α methylserine derivative 14 reveals tolerance for a polar substituent when it is in the R- configuration. As noted above when the amino acid side is acyclic as in L-162,752 (2) α - amino acids are preferred over β -amino acids as in 6. When the amino acid side chain is cyclic as in 15-17, the α -amino analog 15 is surprisingly much less active than the β - and γ -analogs, 16 and 17, respectively.

EC50 for half-maximal release of GH in the rat pituitary cell assay normalized for L-692,429 as 60 nM.

Schoen and Ok^7 have reported that an additional hydroxy or dihydroxy alkyl group in the amino side chain of their benzolactam GH secretagogues dramatically increased GH releasing activity. We, therefore, tried the same changes in the current series of compounds as summarized in Table 2. In analogy with that work, N-alkylation of 6 with a (2R)-hydroxypropyl unit yielded compound 20 with an EC₅₀ of 2.6 nM. This functionality enhanced

the potency of 6 by near 30 fold although the corresponding analog 22 of 2 (L-162,752) was significantly less active than 2.

EC50 for half-maximal release of GH in the rat pituitary cell assay normalized for L-692,429 as 60 nM.

Biological Evaluations

Several studies support the hypothesis that L-162,752 is a mimetic of the hexapeptide GHRP-6. For example, rat pituitary cells stimulated by maximal concentrations of either GHRP-6 or L-162,752 do not respond to further treatment with the other secretagogue, yet remain fully and synergistically responsive to GHRH. Furthermore, the GHRP-6 antagonist His-D-Trp-D-Lys-Trp-D-Phe-Lys-NH₂ antagonizes the effect of L-162,752 on GH release. The *in vitro* specificity of L-162,752 was demonstrated in twenty-four G-protein-linked receptor assays in which IC₅₀ values were >10 µM. Evaluation of L-162,752 in dogs⁵ by intravenous administration showed an increase in serum GH levels at a dose of 0.1 mg/kg. Significant increases of GH levels were observed upon oral dosing of L-162,752 at 2.0 mg/kg (dogs, 2/2) and at 1.0 mg/kg (dogs 1/2). From a comparison of the plasma concentrations of L-162,752 after oral and intravenous administration to rats, its oral bioavailability was estimated to be >40% (Dr. Kwan H. Leung and Dr. Shuet-Hing Lee Chiu, unpublished data). On the other hand, the highly active compound 20 was inactive orally in the dog assay at 1.0 mg/kg as was the more polar compound 21. These results and the good potency of L-162,752 helped establish the importance of the aminoisobutyric acid functionality in our spiroindanylpiperidine growth hormone secretagogues.

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References and Notes:

- (a) Jaffe, C. A.; Ho, P. J.; Demott-Friberg, R.; Bower, C. Y.; Barkan, A. L. J. Clin. Endocrinol. Metab.
 1993, 77, 1641. (b) Ghigo, E.; Awat, E.; Gianotti, L.; Imbimbo, B. P.; Lenaerts, V. Deghenghi, R.;
 Camanni, F. J. Clin. Endrocrinol. Metab. 1994, 78, 693.
- Gertz, B. J.; Barrett, J. S.; Eisenhandler, R.; Krupa, D. A; Wittreich, J. M.; Seibold, J. R.; Scheneider, S. H. J. Clin. Endocrinol. Metab. 1993, 77, 1393.
- 3. Patchett, A. A.; Nargund, R. P.; Tata, J. R.; Chen, M.-H.; Barakat, K. J.; Johnston, D. B. R.; Chen, K. Chan, W. W.-S.; Butler, B.; Hickey, G.; Jacks, T.; Schleim, K.; Pong, S.-S.; Chaung, L.-Y. P.; Chen, H. Y.; Frazier, E.; Leung, K. H.; Chiu, S.-H. L.; Smith, R. G. Proc. Natl. Acad. Sci. 1995, 92, 7001.
- 4. Cheng, K.; Chan, W. W. S.; Barreto, A.; Convey, E. M.; Smith, R. G. Endocrimology 1989, 124, 2791.
- 5. Compounds were administered orally at the appropriate levels in water solution to male and female beagle 1.2-2.5 years old, weighing 8-16 kg. GH levels in plasma were determined by RIA at -5, 0 and +5 and every 10 minutes thereafter for 2 hours.
- 6. Compound 1 was effective i.v. at 0.1 mg/kg (1/1) but inactive at p.o. at 5.0 mg/kg (0/2).
- (a) Schoen, W. R.; Pisano, J. M.; Prendergast, K.; Wyvratt, M. J.; Fisher, M. H.; Cheng, K.; Chan, W. W.-S.; Butler, B.; Smith, R. G.; Ball, R. G. J. Med. Chem. 1994, 37, 897. (b) Schoen W. R.; Ok, D.; DeVita, R. J.; Pisano, J. M.; Hodges, P.; Cheng, K.; Chan, W. W.-S.; Butler, B. S.; Smith, R. G.; Wyvratt, M. J.; Fisher, M. H. Bioorg. Med. Chem. Lett. 1994, 4, 1117.
- 8. Chambers, M. S.; Baker, R.; Billington, D. C.; Knight, A. K.; Middlemiss, D. N.; Wong, E. H. F. J. Med. Chem. 1992, 35, 2033.
- 9. Bodanszky, M. and Bodanszky, A. The Practice of Peptide Synthesis Springer-Verlag, Berlin and Heidelberg, 1984.
- 10. Hanessian, S; Kloss, H. Tetrahedron Lett. 1985, 26, 1261.
- 11. Kuszmann, J.; Tomori, E.; Meerwald I. Carbohydr. Res. 1984, 128, 87.
- 12. pKa's were determined by direct titration in 50% aqueous methanol.
- 13. pKa value of **L-162,752** is 7.9.¹²

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